

## A morphophysiological study in Khamti lahi of rice variety during fluoride stress at seed germination and early seedling stage

CHAU SUNIKTHA CHOWLU<sup>1\*</sup> AND BHABEN CHOWARDHARA<sup>1\*</sup>

Department of Life-Sciences, Faculty of Science and Technology, Arunachal University of Studies, Knowledge City; Namsai-792103, Arunachal Pradesh

Received: August, 2024; Revised accepted: April, 2025

### ABSTRACT

The intense behind conducting these studies was to evaluate the toxicological responses of exposing khamti lahi variety of rice to fluoride stress at germination and earlier seedling stage. The experiment was conducting at department of the Botany at Arunachal University of studies Namsai in January 2023. The surface sterilized seed of uniform size were set of germination over the cotton bed in petri plates, treated with the solution of fluoride of various concentration (0, 0.5, 1mM), prepare from a stock solution NaF (10mM). Results indicated that fluoride could impose significant phyto toxicity during germination and seedling growth during khampti lahi. As compare with the control the germination percentage (55%), germination index (52.2%), seedling vigour index (66.8%), seedling length (33.8%) relative water content (37%) and pigment content was significantly deduced under F stress in the intake 8days old seedling were recorded in khamti lahi variety of rice increasing the dosage of Fluoride.

**Keywords:** Khamti lahi of rice, Fluoride, germination, seedling growth, pigment

### INTRODUCTION

As the primary food crop for 50% of the world's people, rice is farmed on all six continents: Asia, Africa, Australia, Europe, North America, and South America. Though most of the world's rice is grown in Asian countries, India comes in second with 172,580,000 tonnes produced, trailing only China. According to the Food and Agriculture Organization of the United Nations (FAO 2022), Indonesia, Bangladesh, and Vietnam are the top five rice-producing countries in the world. The rice plant belongs to the *Oryza* species of the Poaceae family. Only two of the 24 varieties of the *Oryza* genus are cultivable: *O. sativa* and *O. glaberrima*. There are three further subvarieties of *Oryza sativa*: *indica*, *japonica*, and *javanica*. According to Mahajan *et al.* (2017), India produces rice varieties that are a part of the indica subspecies. The main food crop in India is rice, and the production of this crop represents a major part of the nation's economy. It is a vital food for over 65% of India's population. The Ministry of Agriculture and Farmers Welfare of the Government of India broadly categorized India's rice-growing regions into five primary areas during the 2016–17 fiscal year.

Many unusual and ethnic kinds of rice originate in northeastern India. In many places, only specific kind of rice may be found in that location. Due to the high-yielding output, however, the cultivars currently exclusively grow hybrid and common kinds of rice such basmati, black rice, sticky rice, Ranjit, etc. Thailand lahi is a rare species grown in Arunachal Pradesh that is tall, long grained, and sticky in texture. While Khampti lahi is the primary crop in Namsai district, Arunachal Pradesh, Khampti lahi is a tall plant with long, slender grains that are sticky in texture where Arunachal Pradesh is also known as acidic soil (Sharma *et al.*, 2022; Talukdhar and Beka, 2005). Fluoride is a form of fluorine that is an anion. The halogen family of chemicals includes fluoride. According to Greenwood and Earnshaw, (2012), the term "fluoride" can refer to both organic and inorganic fluorine compounds. Fluoride does not occur in nature in its elemental form due to its high reactivity. The oxidative state of the fluoride ion is -1, ranking it as the 13th most prevalent element in the crust of the planet. Numerous environmental, medicinal, and nutritional samples contain it. It is most usually found in the form of chemical compounds like sodium fluoride or hydrogen fluoride in the minerals fluorospar, fluorapatite, topaz, and cryolite (Shashank and Balaaji 2011).

\*Corresponding authorship: bhaben1986@gmail.com, chowlusunik@gmail.com

According to Agalakova and Gusev (2012), fluorides are a prevalent, hazardous, non-biodegradable non-metal pollutant. Fluoride-induced non-metal-induced soil contamination is one of the world's largest problems (Chaudhary and Khan, 2014). Fluoride is necessary for normal plant growth in small doses, but in higher quantities, it can potentially harm both plants and the environment. The manufacturing of aluminum and the mining, processing, and use of phosphate rock as fertilizer for agriculture are the two main human activities that emit fluorides into the environment. Other manufacturing procedures (such as the production of steel, copper, nickel, glass, brick, ceramic, glues, and adhesives) as well as the burning of coal, which contains fluoride impurities, are other sources of fluoride. Once it is in a stable state, fluoride stays in the environment for a reasonable amount of time until it transforms into another chemical or degrades as a result of radiation. The origin and/or absence of clay, the level of organic carbon, the pH, and the soil are the main contributors to fluoride in soils. Additionally, it has been discovered that the fluoride's bioavailability to plants is influenced by the water-soluble fluoride found in saline soils (Al Sabti *et al.*, 2023).

Fluoride generally enters vegetation through two different channels. According to Kamaluddin and Zwiazek, (2003), fluoride initially penetrates the cell walls through stomatal diffusion, permeates them, and then moves to the margins and tips, which are the areas that experience the maximum evaporation. The second channel employs passive diffusion to allow soil and water to reach plant roots. The apoplastic and symplastic xylem channels are subsequently used to transfer fluoride to the shoots in a unidirectional distal migration (Pant *et al.* 2008). Hydrogen fluoride (HF), which diffuses non-ionically, also transports fluoride through cellular membranes. Because the tiny neutral particle of HF enters cell membranes significantly more quickly than a dissociated fluoride ion (F<sup>-</sup>), there is a more pronounced intracellular intake. Comparing HF to F<sup>-</sup>, membrane leakage is increased by five to seven orders of magnitude (Pant *et al.* 2008). Fluoride toxicity has an impact on a variety of morphological, physiological, and biochemical processes, including seed germination parameters, growth, development, mineral nutritional status, photosynthesis, respiration,

metabolic activity, yield and yield characteristics, and others (Sahariya *et al.* 2021). Researchers have discovered that fluoride-treated plants have significantly lower growth and related parameters than control plants, including seedling germination %, roots and shoot length, plant height, and fresh and dry biomass (Singh *et al.*, 2013). In this article, mainly focus the effect of fluoride in Local Khamti lai rice variety where the Arunachal Pradesh becomes acidic soil.

## MATERIAL AND METHODS

The local variety seeds of *Oryza sativa* L., also known as Khamti lai, were collected from Namsai's local farmers. After being surface sterilized in a 1% (w/v) NaOCl solution for 15 minutes, the seeds were repeatedly rinsed with distilled water. After being sterilized and sized uniformly, the seeds were treated with a sodium chloride solution derived from a stock solution (1.0M) and placed on a cotton bed in a petri plate for germination. Four NaF dosages (0, 0.5 and 1.0 mM) were examined using a randomized approach in three replicates. Each glass petri plate held twenty seeds, to which 10.0 ml of a solution (distilled water or sodium fluoride) was added. The following protocols were used to measure the final germination percentage (FGP), germination index (GI), germination energy (GE), and finally the seedling vigour index (SVI) and seedling length: Li. (2008); Abdul-Baki and Anderson (1973); and Moulick *et al.* (2016). A few specific biochemical parameters, like the amount of carotenoid and chlorophyll, were measured using the Arnon (1949) methodology. The software SPSS 21 (Window version) was used to analyze the data using correlation analysis, mean (n=3), and standard error (mean SE) formats.

## RESULTS

Seed germination is an important stage in the life cycle of any plant, and it is greatly influenced by a range of environmental conditions (Chowardhara *et al.* 2020).

### Effect of Sodium fluoride on germination of khampti lahi

Under the stress condition NaF, the germination% rate was significantly ( $p < 0.001$ )

reduced at 1mM (55%) or discriminated as the days increases with compare to control respectively (Table 1). As a result, it also effects on germination index where the concentration increases the germination index also reduced (25 and 55%) at 0.25mM and 1mM as compare with control respectively Table 1. The germination energy also influences by NaF as compared with control where 55% was reduced at 1mM significantly as well as also Relative germination rate also effected as the concentration decreases to 55% and 25% at

0.5mM and 1mM as compare with control (Table 1). The Relative metal injury index was also measure in khampti lahi under the NaF stress condition as a result the relative metal injury index was significantly hampered ( $P < 0.001$ ).

Under NaF stress condition, the moisture content as well as Relative water content was significantly drop in seedling plants as compared untreated plants (Table 1) Along with the Seedling vigour index was also significantly was reduced 67% at higher concentration.

Table 1: Consequences of F toxicity on germination, seedling growth, germination energy, relative injury index, germination index and seedling vigor index (intact seedlings) at 8 DAS

NaF Conc. (mM)	FGP	GE	GI	RGR	RSIR	SVI
0	100±0	10±0	12.5±0	1±0	1	1348.77±0.77
50	75± 1.0***	7.5±0.5*	9.37±0.62*	0.75±0.058	0.25±0.10***	832.38±0.38***
100	45± 1.0***	4.5± 0.5**	5.93±0.31**	0.45±0.18**	0.55±0.22**	446.45±0.45***

#### Effect of NaF on Morphomatric attribute

Under the NaF stress condition, the seedling length was significantly reduced as a concentration of fluoride in khampti lahi variety due to higher dosage of fluoride which inhibit the cell division. The seedling length at high

concentration was significantly decreases (34%) khampti lahi seedling plants as compare to untreated plants (Table 2). Similar result also observed in biomass that is Fresh weight (59%) and Dry weight (78%) was significantly reduced as the dosage increases (at 1mM NaF) whereas compare to control plants.

Table 2: Consequences of NaCl toxicity on seedling growth, fresh weight, turgor weight, dry weight and relative water content (intact seedlings) at 7 DAS (days after sowing)

NaF Conc. (mM)	Seedling length (cm)	FW (gm)	DW (gm)	RWC
0	13.66±0.18	0.88±0.01	0.66±0.01	0.145±0.005
50	10.43±0.03*	0.59±0.01***	0.435±0.02**	0.125±0.005 <sup>ns</sup>
100	8.38±0.54**	0.375±0.005***	0.145±0.005***	0.085±0.005*

#### Effect of NaF on Chlorophyll pigment

Chl a (90%), Chl b (87%), total chl (73%), Ratio of Chl a and chl b (1.03) was significantly

deduced in khampti lahi under NaF stress condition (Table 3) as compared with control due to the damage pigment structure damage.

Table 3: Consequences of NaCl toxicity on Chlorophyll a, Chlorophyll b, total chlorophyll, Carotenoid and Chlorophyll a and b ratio (intact seedlings) at 7 DAS (days after sowing)

NaF Conc. (mM)	Chl a (mg g <sup>-1</sup> Fw)	Chl b (mg g <sup>-1</sup> Fw)	Total Chl (mg g <sup>-1</sup> Fw)	Carotenoid (mg g <sup>-1</sup> Fw)
0	23.15±0.35	17.72±0.47	41.55±0.45	13.69±0.40
50	5.94±0.06***	10.89±0.30**	16.87±0.12***	3.89±0.11***
100	2.28±0.07***	2.04±0.06***	4.55±0.05***	3.86±0.02***

#### Membrane Injury Index

Membrane Injury Index meaning leakage of ions from seedling plants. The result showed

that the gradually enhanced the concentration of 0.5mM and 1mM as a consequence of control seedling plants.

Table 3: Correlation coefficients among GP%, GI, GE, RGR, RMIR, SVI, SL, FW, DW TW, RWC, Chl a, Chl b, total chl and carotenoids

Parameters	GP%	GI	GE	RGR	RMIR	SVI	SL	FW	DW	TW	RWC	Chl a	Chl b	Total Chl	carotenoids
GP%	1	0.99** *	0.99** *	0.99***	-0.99***	0.98***	0.94**	0.06*	0.99***	0.502 <sup>ns</sup>	0.99** *	0.956*	0.903*	0.962**	0.81*
GI		1	0.967* *	0.995** *	- 0.995** *	0.993***	0.961**	0.986***	0.997* **	0.567 <sup>ns</sup>	0.946* **	0.933** *	0.995* **	0.978** *	0.850*
GE			1	0.987** *	- 0.985** *	0.967**	0.914*	0.925**	0.454 <sup>n</sup> s	0.984** *	0.935* *	0.871* **	0.987* **	0.939** *	0.785ns
RGR				1	-0.99***	0.990***	0.49**	0.969***	0.521 <sup>n</sup> s	1.00***	0.952* *	0.913* *	1.0***	0.969** *	0.826*
RMI					1	-0.991***	-0.58**	-0.972**	- 0.530 <sup>n</sup> s	- 0.999** *	- 0.950* *	- 0.917** *	1.00** *	-0.71***	-0.833*s
SVI						1	0.977***	0.991***	0.636 <sup>n</sup> s	0.0991* **	0.961* **	0.991** *	0.991* **	0.911* *	0.961**
SL							1	0.981***	0.704 <sup>n</sup> s	0.950**	0.865* *	0.970** *	0.950* **	0.983** *	0.925**
FW								1	0.276 <sup>n</sup> s	0.733*	0.867* *	0.867* *	0.733* *	0.333 <sup>ns</sup>	0.333 <sup>ns</sup>
DW									1	0.867* *	0.867* *	0.867* *	0.733* *	0.333 <sup>ns</sup>	0.333 <sup>ns</sup>
TW										1	0.247 <sup>n</sup> s	0.138 <sup>ns</sup>	0.414 <sup>n</sup> s	0.414 <sup>ns</sup>	0.828*
RWC											1	0.733* *	0.733* *	0.600 <sup>ns</sup>	0.200ns
Chl a												1	0.943* *	0.657 <sup>ns</sup>	0.429 <sup>ns</sup>
Chl b													1	0.943**	0.657
Total Chl														1	0.714ns
Carotenoids															1

\* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ 

## Correlation analysis

The result showed analysis among Germination %, Germination Index, Germination Energy, Relative germination rate, Seedling vigour index, Seedling length, fresh weight, dry weight, turgid weight, Relative water content chlorophyll *a*, *b* total chl, Relative metal injury most of the parameters are positive correlation ship except RMIR with GP, GR, GE, RGR, SVI, SL, FW, DW, TW, RWC, chl *a*, chl *b*, total chl and carotenoid (Table 4) i.e., the increase of the concentration of the NaF also effect on the others parameters also.

## DISCUSSION

Phytase, an enzyme, breaks down phytin during germination to give the developing seedling inorganic phosphate. Fluoride suppresses the dephosphorylation of phytin molecules in tissues, which reduces seedling root growth during germination. Fluoride also inhibits the phytase enzyme. For the metabolism of RNA, orthophosphate sources produced from

phytin are necessary. Lack of phytin-derived orthophosphates is one of the factors that could limit the growth of seedlings treated with fluoride. Increases in fluoride concentration were found to reduce seed germination in *Triticum aestivum*, *Zea mays*, *Helianthus annuus*, and *Vicia faba* (Shadad *et al.* 1989), as well as *Cicer arietium* (L), (Kumar *et al.* 2017 and Pelc *et al.* 2020), *Solanum lycopersicum* (Ahmad *et al.* 2018), *Cajanus cajan* L. (Yadu *et al.* 2018), cv Anuradha (Datta *et al.* 2012), *Raphanus sativus* L. (Singh *et al.* 2013), Fenugreek (Burgohain and Chowardhara, 2022). According to Shabl *et al.* (2006), increasing the concentration of NaF considerably reduces *Cyamopsis tetragonoloba* seed germination, seedling growth, and biomass when compared to the control. *Oryza sativa* has also been linked to similar discoveries (Gupta *et al.* 2009; Banerjee and Roychoudhury, 2019a; Singh and Roychoudhury, 2020; Banerjee and Roychoudhury, 2021a). One of the most crucial processes in plants is photosynthesis. The photosynthesis is influenced by a variety of circumstances, including heavy metal impact on plants. Different air contaminants may impair a number of metabolic pathways and mechanisms

as well as the production of agricultural crops (Kumar *et al.* 2017). Plant response to pollution is influenced by the chemical element's toxicity, the length of exposure, and the species' sensitivity (Oguchi *et al.* 2005). Due to its electromotivity, electronegativity, and significant phytotoxic potential, F stands out among the pollutants. Above all of these factors, it has the capacity to preferentially enter through the stomata (Franzaring *et al.*, 2007). The cells and tissues in the leaves suffered ultrastructural and structural damage as a result of the F buildup. Plant stomatal conductance and gas exchange will be significantly impacted by the damage to cells and tissues. The epidermis and stomata of young coffee (*Coffea arabica*) and orange (*Citrus X sinensis*) plants are harmed when exposed to hydrogen fluoride (HF) in a semi-open mist chamber, according to Mesquita *et al.* (2011). It might lead to a breakdown in the mechanisms that regulate stomatal opening and closing. F decreases the activity of several enzymes in the chloroplasts, including ATP synthase, ribulose biphosphate carboxylase-oxygenase (RuBisCo), and sucrose synthase. Fluoride greatly increased in concentration in plants impeded photosynthesis as well. F reduces the production of chlorophyll, destroys chloroplasts, and slows down the Hill's response to prevent photosynthesis. A decrease in chlorophyll content also has an adverse effect on plant photosynthetic systems. Finally, this led to a decrease in CO<sub>2</sub> generation and assimilation (Domingues *et al.* 2011). The photosynthetic electron transport pathway in plant thylakoid membranes was examined after F exposure. The electron transport rate of PSII is inhibited as F builds up, and then PS-I's electron transport rate rises as a result. This result suggested state changes as a potential mechanism for F toxicity. F treatment at 190 ppm reduces photosynthetic pigments in plants. *Salicornia brachiata* Roxb showed comparable results in Reddy and Kaur (2008) study. Plants grown in F-polluted soil showed decreased photosynthetic capacity, Chl-*a* and Chl-*b* concentrations, total chlorophyll, carotenoids, and leaf area (Kumar and Rao, 2008; Ram *et al.* 2014). According to Gupta *et*

*al.* (2009), F may have hindered chlorophyll production, lowering plant chlorophyll concentration. The amount and activity of the enzyme chlorophyllase, which breaks down chlorophyll, are anticipated to rise when F accumulation (Ram *et al.*, 2014). When plants were grown on soil contaminated with F in the semi-arid region, similar effects were seen (Baunthiyal and Sharma, 2014). Fluoride concentrations above a particular threshold have an adverse effect on photosynthesis, which leads to a decrease in the formation of dry matter (Zouari *et al.*, 2016). SVI reduced in higher concentration than to control by (33% and 66%) in 0.5mM and 1mM as a same have been studied in *Trigonella foenum-graecum* at 7 DAS that it significantly decreased similar to the khampti lahi by (37.6%, 60.6% and 80%) of 2,4,6mM concentration of fluoride.

In other parameters stress has also been found as same like in Cadmium and Zinc on Indian mustard (*Brassica juncea*) (Chowardhara *et al.* 2020; Chowardhara *et al.* 2019). Membrane injury index is almost equal in all the concentration which shows that in control injury has been hampered MII has also studied in other stress like in Auminium which shows gradually increased from control to higher concentration (Panda *et al.* 2003).

## CONCLUSION

The findings suggested that fluoride may cause serious phytotoxicity during khampti lahi seedling development and germination. Under F stress in the input, there was a substantial decrease in the germination percentage (55%), germination index (52.2%), seedling vigour index (67%), seedling length (34%), relative water content (37%), and pigment content as compared to the control. In the Khampti Lahi type of rice, 8-day-old seedlings were seen when the fluoride dosage was increased. Since khampti lahi is a native rice plant that is grown in a specific area of the Namsai district in Arunachal Pradesh, no research has been done on it under abiotic stress conditions up to this point.

## REFERENCES

- Agalakova, N.I., and Gusev, G.P. (2012) Fluoride induces oxidative stress and ATP depletion in the rat erythrocytes in vitro. *Environmental toxicology and pharmacology*, **34**(2), 334-337.
- Ahmad, M.A., Bibi, H., Munir, I., Ahmad, M.N.,

- Zia, A., Mustafa, G., Ullah, I. and Khan, I., (2018) Fluoride toxicity and its effect on two varieties of *Solanum lycopersicum*. *Fluoride*, **51**(3), pp.267-277.
- Al Sabti, B., Samayamanthula, D. R., Dashti, F. M., and Sabarathinam, C. (2023) Fluoride in Groundwater: Distribution, Sources, Processes, Analysis, and Treatment Techniques: A Review. *Hydrogeochemistry of Aquatic Ecosystems*, 1-31.
- Banerjee, A., and Roychoudhury, A. (2019a) Differential regulation of defence pathways in aromatic and non-aromatic indica rice cultivars towards fluoride toxicity. *Plant Cell Reports*, **38**, 1217-1233.
- Banerjee, A., and Roychoudhury, A. (2021) Fluoride-induced toxicity is ameliorated in a susceptible indica rice cultivar by exogenous application of the nitric oxide donor, sodium nitroprusside. *Vegetos*, **34**(3), 568-580.
- Baunthiyal, M., and Sharma, V. (2014) Response of three semi-arid plant species to fluoride; consequences for chlorophyll florescence. *International journal of phytoremediation*, **16**(4), 397-414.
- Burgohain, U and Chowardhara, B. (2022) Fluoride induces morphological change in Fenugreek (*Trigonella foenum-graecum* L) during germination and early seedling. *Annals of Plant and Soil Research*, **24** (4), 596-600.
- Chaudhary, K., and Khan, S. (2014) Effect of plant growth promoting rhizobacteria (PGPR) on plant growth and flouride (F) uptake by F hyperaccumulator plant *Prosopis juliflora*. *International Journal of Recent Scientific Research*, **5**(11), 1995-9.
- Chowardhara, B., Borgohain, P., Saha, B., Awasthi, J. P., and Panda, S. K. (2020) Differential oxidative stress responses in *Brassica juncea* (L.) Czern and Coss cultivars induced by cadmium at germination and early seedling stage. *Acta Physiologiae Plantarum*, **42**, 1-12.
- Chowardhara, B., Borgohain, P., Saha, B., Awasthi, J. P., Moulick, D., and Panda, S. (2019) Assessment of phytotoxicity of zinc on Indian mustard (*Brassica juncea*) varieties during germination and early seedling growth. *Annals of Plant and Soil Research*, **21**(3), 239-244.
- Datta, J.K., Maitra, A., Mondal, N.K., and Banerjee, A. (2012) Studies on the impact of fluoride toxicity on germination and seedling growth of gram seed (*Cicer arietinum* L. cv. Anuradha). *Journal of Stress Physiology & Biochemistry*, **8**(1), 194-202.
- Domingues, R.R., Mesquita, G.L., Cantarella, H., and Mattos Júnior, D.D. (2011) Suscetibilidade do capim-colonião e de cultivares de milho ao flúor. *Bragantia*, **70**, 729-736.
- Franzaring, J., Hrenn, H., Schumm, C., Klumpp, A., and Fangmeier, A. (2006) Environmental monitoring of fluoride emissions using precipitation, dust, plant and soil samples. *Environmental pollution*, **144**(1), 158-165.
- Greenwood, N.N., and Earnshaw, A. (2012) *Chemistry of the Elements*. Elsevier.
- Gupta, S., Banerjee, S., and Mondal, S. (2009) Phytotoxicity of fluoride in the germination of paddy (*Oryza sativa*) and its effect on the physiology and biochemistry of germinated seedlings. *Fluoride*, **42**(2), 142.
- Joint, F. A. O., World Health Organization, and WHO Expert Committee on Food Additives. (2022) Evaluation of certain contaminants in food: ninetieth report of the Joint FAO/WHO Expert Committee on Food Additives.
- Kamaluddin, M., and Zwiazek, J. J. (2003) Fluoride inhibits root water transport and affects leaf expansion and gas exchange in aspen (*Populus tremuloides*) seedlings. *Physiologia Plantarum*, **117**(3), 368-375.
- Kumar, K. A., and Rao, A. V. B. (2008) Physiological responses to fluoride in two cultivars of mulberry. *World J Agric Sci*, **4**(4), 463-466.
- Kumar, K., Giri, A., Vivek, P., Kalaiyarasan, T., and Kumar, B. (2017) Effects of fluoride on respiration and photosynthesis in plants: an overview. *Annals of Environmental Science and Toxicology*,

- 2(1), 043-047.
- Mahajan, G., Kumar, V., and Chauhan, B. S. (2017) Rice production in India. *Rice production worldwide*, 53-91.
- Mesquita, G. L., Tanaka, F. A. O., Cantarella, H., and Mattos, D. (2011) Atmospheric absorption of fluoride by cultivated species. Leaf structural changes and plant growth. *Water, Air, & Soil Pollution*, **219**, 143-156.
- OGUCHI, R., HIKOSAKA, K., and Hirose, T. (2005) Leaf anatomy as a constraint for photosynthetic acclimation: differential responses in leaf anatomy to increasing growth irradiance among three deciduous trees. *Plant, Cell & Environment*, **28**(7), 916-927.
- Pant, S., Pant, P., and Bhiravamurthy, P. V. (2008) Effects of fluoride on early root and shoot growth of typical crop plants of India. *Fluoride*, **41**(1), 57.
- Pelc, J., Śnioszek, M., Wróbel, J., and Telesiński, A. (2020) Effect of fluoride on germination, early growth and antioxidant enzymes activity of three winter wheat (*Triticum aestivum* L.) cultivars. *Applied Sciences*, **10**(19), 6971.
- Ram, A., Verma, P., and Gadi, B. R. (2014) Effect of fluoride and salicylic acid on seedling growth and biochemical parameters of watermelon (*Citrullus lanatus*). *Fluoride*, **47**(1), 49-55.
- Reddy, M. P., and Kaur, M. (2008) Sodium fluoride induced growth and metabolic changes in *Salicornia brachiata* Roxb. *Water, Air, and Soil Pollution*, **188**, 171-179.
- Sabal, D., Khan, T. L., and Saxena, R. (2006) Effect of sodium fluoride on cluster bean (*Cyamopsis tetragonoloba*) seed germination and seedling growth. *Fluoride*, **39**(3), 228.
- Sahariya, A., Bharadwaj, C., Emmanuel, I., and Alam, A. (2021) Phytochemical profiling and GCMS analysis of two different varieties of barley (*Hordeum vulgare* L.) under fluoride stress. *The Scientific Temper*, **12**(1&2).
- Shadad, M.E. (1989) Proximate composition, tannin content and protein digestibility of sorghum cultivars grown in Sudan. *Sci. (Agric.) Thesis, University of Khartoum, Sudan*.
- Sharma, A., Monlai, S., Devadas, V. S., Libang, J., and Buri, C. (2019) Effect of climatic factors on seedling growth of local and exotic rice varieties. *Journal of Pharmacognosy and Phytochemistry*, **8**(3), 1973-1980.
- Shashank, R., and Balaaji, V. B. (2011) Fluoride Estimation and Reduction in Indian Water Samples using Optimum Alumina, pH and Time of Operation. *International Journal of Chemical Engineering and Applications*, **2**(1), 38.
- Singh, A., and Roychoudhury, A. (2020) Silicon-regulated antioxidant and osmolyte defense and methylglyoxal detoxification functions co-ordinately in attenuating fluoride toxicity and conferring protection to rice seedlings. *Plant Physiology and Biochemistry*, **154**, 758-769.
- Singh, S., Singh, J., and Singh, N. (2013) Studies on the impact of fluoride toxicity on growth parameters of *Raphanus sativus* L. *Indian Journal of Scientific Research*, **4**(1), 61-63.
- Talukdar, K.C., and Beka, B.C. (2005) Cultivation of summer rice in the flood plains of Assam—An assessment of Economic Potential on marginal and small farms. *Agricultural Economics Research Review*, **18**(1), 21-38.
- Yadu, B., Chandrakar, V., Meena, R. K., Poddar, A., and Keshavkant, S. (2018). Spermidine and melatonin attenuate fluoride toxicity by regulating gene expression of antioxidants in *Cajanus cajan* L. *Journal of plant growth regulation*, **37**, 1113-1126.
- Zouari, M., Elloumi, N., Bellassoued, K., Ahmed, C.B., Krayem, M., Delmail, D., Elfeki, A., Rouina, B.B., Abdallah, F.B. and Labrousse, P., (2017) Enzymatic antioxidant responses and mineral status in roots and leaves of olive plants subjected to fluoride stress. *South African Journal of Botany*, **111**, pp.44-49.